Beckwith-Wiedemann Syndrome

Authors: Doctor Christine Gicquel¹, Doctor Sylvie Rossignol, Professor Yves Le Bouc Creation Date: September 2001 Updated: March 2005

Scientific Editor: Professor Didier Lacombe

¹Laboratoire d'explorations fonctionnelles, Hôpital Trousseau, 26 Avenue du Docteur Arnold Netter, 75571 Paris Cedex 12, France. <u>christine.gicquel@trs.ap-hop-paris.fr</u>

Abstract Keywords Disease name and synonyms Definition Diagnosis criteria Differential diagnosis Prevalence Etiology Genotype/phenotype correlations Diagnostic methods Management of BWS patients Genetic counseling Antenatal diagnosis Beckwith-Wiedemann syndrome and assisted reproductive technology References

Abstract

Beckwith-Wiedemann Syndrome (BWS; OMIM 130650) is an overgrowth disorder characterized by macrosomia, macroglossia, organomegaly and developmental abnormalities (in particular abdominal wall defects with exomphalos). Its incidence is estimated to be 1 per 13,700 live births. BWS patients are prone to the development of embryonal tumors (most commonly Wilms' tumor or nephroblastoma). BWS is a multigenetic disorder caused by dysregulation of gene expression in the imprinted 11p15 chromosomal region. Various 11p15 defects have been implicated and epigenetic defects account for about two thirds of cases. The management of patients with BWS involves the surgical cure of exomphalos and monitoring of hypoglycemia in the neonatal period. It also involves the treatment of macroglossia and the screening for embryonal tumor that can be facilitated by genotyping. A recent series of reports suggested that assisted reproductive technology (ART) may increase the risk of imprinting disorders, and BWS in particular.

Keywords

Overgrowth syndrome, Beckwith-Wiedemann syndrome, 11p15 region, *IGF2* gene, *H19* gene, *KCNQ1OT1* gene, *CDKN1C* gene, embryonal tumor, imprinting, methylation, epigenetic change, assisted reproductive technology

Disease name and synonyms

Beckwith-Wiedemann syndrome (BWS) Exomphalos-Macroglossia-Gigantism syndrome (EMG syndrome)

Definition

BWS is an overgrowth disorder involving developmental abnormalities. This multigenic disorder is caused by dysregulation of the expression of imprinted genes in the 11p15 chromosomal region.

Gicquel C., Rossignol S., Le Bouc Y. Beckwith-Wiedemann Syndrome. Orphanet encyclopedia. March 2005. http://www.orpha.net/data/patho/GB/uk-BWS05.pdf

Diagnosis criteria

(for references see: Wiedemann HR, 1964; Pettenati MJ *et al.*, 1986; Elliott M *et al.*, 1994; DeBaun MR *et al.*, 1998)

The phenotypic expression of BWS is variable and diagnosis is still based on clinical signs. Recent improvements in the molecular diagnosis of BWS and other overgrowth disorders suggest that in the next few years, BWS will be defined molecularly.

It is generally accepted that diagnosis of BWS requires at least 3 clinical findings including at least 2 major findings:

Major clinical findings

- Macroglossia (present in more than 95% of patients).

- Macrosomia or overgrowth, defined as preand-/-or postnatal growth greater than the 97th percentile (present in about 80% of patients). The trend to increased size continues through early childhood but becomes less dramatic with increasing age.

- Abdominal wall defects (exomphalos, umbilical hernia, diastasis recti; 65% of patients).

- Organomegaly involving principally abdominal organs: kidneys, liver, spleen, pancreas and adrenal glands (present in 50% of patients).

Minor clinical findings

- Hypoglycemia in the neonatal period (occurs in about 40% of patients), mostly mild and transient.

- Renal abnormalities: malformations, medullary dysplasia.

- Ear creases and pits (30% of patients).
- Facial nevus flammeus (30% of patients).
- Hemihyperplasia (30-35% of patients).

- Embryonal tumors: about 7.5% of BWS patients develop tumors (Wilms' tumor, neuroblastoma, adrenal carcinoma, hepatoblastoma, rhabdomyosarcoma), most commonly in the first 6 years of life.

- Polyhydramnios.

Differential diagnosis

Clinically, BWS must be distinguished from other overgrowth syndromes:

- <u>Simpson-Golabi-Behmel syndrome</u> (OMIM 312870) is an X-linked disease caused by mutations in the gene encoding glypican-3, an extracellular proteoglycan known to play an important role in growth control in embryonal mesoderm tissues, in which it is selectively expressed. This proteoglycan binds IGF2, reducing its availability to the type 1 IGF-receptor (Pilia G *et al.*, 1996; Neri G *et al.*, 1998).

- <u>Perlman syndrome</u> (OMIM 267000), which is often more severe, has a high perinatal mortality rate. Its pathogenesis is still unknown (Grundy RG *et al.*, 1992; Henneveld HT *et al.*, 1999). - <u>Sotos syndrome</u> (OMIM 117550) is caused by mutations in the gene encoding NSD1 and should be considered for differential diagnosis because of the clinical overlap with BWS (Baujat G *et al.*, 2004).

BWS must also be recognized in its incomplete forms (Sotelo-Avila C *et al.*, 1980) and in related forms, such as non-syndromic IGF2 overgrowth disorder (Morison IM *et al.*, 1996). Some cases of isolated hemihyperplasia are also related to abnormalities in the 11p15 region and are associated with a risk of tumor (Hoyme HE *et al.*, 1998).

Prevalence

The incidence of BWS (1 per 13,700 livebirths) has been reported in only one study (Thorburn MJ *et al.*, 1970) and is probably underestimated.

Etiology

BWS is caused by imprinting errors in the 11p15 chromosomal region (Maher ER et al., 2000; Reik W et al., 2001) This region includes genes encoding growth factors and tumor suppressor The paternally expressed genes. genes (maternally imprinted) have growth enhancing activity and the maternally expressed genes (paternally imprinted) have growth suppressing activity. The 11p15 region is organized into two domains: a telomeric domain including the IGF2 and H19 genes and a centromeric domain including the CDKN1C (Cyclin Dependant Kinase Inhibitor 1C), KCNQ1 (Potassium voltage-gated channel, subfamily Q, member 1) and KCNQ10T1 (KCNQ1-Overlapping transcript 1) genes. Each domain is controlled by its own imprinting center (IC1 and IC2 for the telomeric and the centromeric domains, respectively) (Reik W et al., 2001).

BWS is a multigenic disorder involving various molecular abnormalities in the 11p15 region (Engel JR *et al.*, 2000; Bliek J *et al.*, 2001; Gaston V *et al.*, 2001; Weksberg R *et al.*, 2001; DeBaun MR *et al.*, 2002).

- Cytogenetic abnormalities account for 1-2% of cases and consist of maternally inherited translocations or inversions and trisomy with paternal duplication.
- The genetic abnormalities described in BWS are:

- 11p15 paternal uniparental disomy (UPD) of both the centromeric and telomeric domains. In paternal UPD, the maternal allele is lost and the paternal allele is duplicated. This occurs in approximately 20% of cases.

- Mutations in the *CDKN1C* gene (also known as p57KIP2) encoding a maternally expressed cellcycle regulator, found in about 5% of patients (Lam WW *et al.*, 1999). Patients with *CDKN1C* gene mutations have a typical BWS phenotype, with a very high frequency of exomphalos.

Gicquel C., Rossignol S., Le Bouc Y. Beckwith-Wiedemann Syndrome. Orphanet encyclopedia. March 2005. http://www.orpha.net/data/patho/GB/uk-BWS05.pdf Mutations in the *CDKN1C* gene are much more frequent in familial BWS and about 60% of familial BWS cases are caused by mutation of the *CDKN1C* gene.

• The epigenetic abnormalities described in BWS are:

- Hypermethylation of the *H19* gene, a maternally expressed untranslated RNA with tumor suppressor function, found in 10% of cases.

- Demethylation of *KvDMR1*, a differentially methylated region at the 5' end of the *KCNQ1OT1* gene, involved in 55-60% of patients. The *KCNQ1OT1* gene (also known as *LIT1* or *KvLQT1-AS*) encodes an antisense transcript within intron 10 of the *KCNQ1* gene and is normally expressed from the paternal allele (Lee MP *et al.*, 1999; Smilinich NJ *et al.*, 1999). This gene may be involved in regulating imprinting of the centromeric domain (Cleary MA *et al.*, 2001; Fitzpatrick GV *et al.*, 2002).

- It was recently shown that microdeletions within *IC1 (H19 DMR)* (Sparago A *et al.*, 2004) or *IC2* (Niemitz EL *et al.*, 2004) account for a low percentage of BWS cases with hypermethylation of *H19* or demethylation of *KCNQ10T1*. However, the exact frequency of these microdeletions is still unknown.

Genotype/phenotype correlations

The clinical expression of BWS may differ between patients with similar molecular abnormalities, due to the mosaicism distribution for most known molecular defects. This is well-illustrated for 11p15 UPD (Itoh N *et al.*, 2000).

Other genotype/phenotype correlations have also been observed, providing evidence that aspects of the BWS phenotype may be correlated with the involvement of specific imprinted genes (Engel JR *et al.*, 2000; Gaston V *et al.*, 2001; DeBaun MR *et al.*, 2002). Indeed, exomphalos is more frequent in patients with a defect of the centromeric domain (demethylation of *KCNQ10T1* and mutation of *CDKN1C*). Hemihyperplasia and organomegaly are more frequent in patients with hypermethylation of *H19* or 11p15 UPD.

About 7.5 to 10% of BWS patients will develop a tumor. Wilms' tumor is the most common tumor found in patients with BWS (60% of all tumors), but other solid childhood tumors also are found. Previous studies (Engel JR et al., 2000; Bliek J et al., 2001; Gaston V et al., 2001; Weksberg R et al., 2001; DeBaun MR et al., 2002) have clearly shown that 11p15 UPD and H19 hypermethylation are strongly associated with tumor risk in BWS patients. Wilms' tumor is only found in BWS patients with molecular lesions in the telomeric domain and is the only type of tumor found in patients with H19 hypermethylation (Bliek J et al., 2004). Patients

with molecular lesions in the centromeric domain (demethylation of *KCNQ10T1* or mutation of *CDKN1C*) have a low risk of tumor and these patients develop a different spectrum of tumors, including hepatoblastoma, rhabdomyosarcoma and gonadoblastoma (Weksberg R *et al.*, 2001, Bliek J *et al.*, 2004). The only tumor reported in patients with mutation of *CDKN1C* is neuroblastoma (Lee MP *et al.*, 1997; Gaston V *et al.*, 2001).

Diagnostic methods

Careful cytogenetic analysis of the 11p15 region and fluorescent *in situ* hybridization (FISH) can be used to recognize the rare translocations, inversions and trisomies.

Molecular diagnosis is difficult, mostly because of the large spectrum of genetic and epigenetic abnormalities. Molecular tests must differentiate the various abnormalities in the 11p15 region: patients with 11p15 paternal UPD, patients with hypermethylation of the *H19* gene, patients with demethylation of the *KCNQ10T1* gene and patients with a mutation in the *CDKN1C* gene.

As demethylation of the *KCNQ10T1* gene is never associated with abnormal methylation of the *H19* gene except in patients with 11p15 paternal UPD, analysis of the methylation status of both the *KCNQ10T1* and *H19* genes leads to the diagnosis of more than 90% of 11p15 defects:

- Isolated demethylation of the *KCNQ10T1* gene.

- Isolated hypermethylation of the H19 gene.

It is not yet possible to determine precisely the percentage of cases with epigenetic defects displaying a microdeletion of *IC1* or *IC2*.

Hypermethylation of the *H19* gene associated with demethylation of the *KCNQ10T1* gene is indicative of 11p15 paternal UPD, which should be confirmed by analysis of markers of the 11p15 region and of parental DNA. 11p15 paternal UPD always occurs as mosaicism and, because tissue distribution of mosaicism is variable, tissue from a second source (such as fibroblasts) may be helpful.

If the methylation status of the *KCNQ10T1* and *H19* genes is normal, then sequencing of the *CDKN1C* gene is indicated, particularly in patients with exomphalos and/or a family history of BWS.

Management of BWS patients

Neonates with exomphalos should undergo abdominal wall repair soon after birth.

Hypoglycemia during the first few days of life can be anticipated by monitoring glycemia in newborns with BWS every six hours for the first few days. Serious neurological sequelae can therefore be prevented.

Gicquel C., Rossignol S., Le Bouc Y. Beckwith-Wiedemann Syndrome. Orphanet encyclopedia. March 2005. http://www.orpha.net/data/patho/GB/uk-BWS05.pdf

Macroglossia should be treated by a maxofacial surgical team.

Assessing tumor risk is the main difficulty in patients with BWS. Between 7.5% and 10% of all BWS patients will develop a tumor, mostly during the first 6 years of life. The severity of the phenotype, hemihyperplasia and organomegaly (of the kidneys in particular) have been shown to be associated with an increase in the relative risk of cancer (Schneid S et al., 1997; Beckwith JB, 1998; DeBaun MR et al., 1998; Gaston V et al., 2001) but none of these clinical features can identify with certainty patients likely to develop tumors. Based on molecular analysis, it is now possible to discriminate between BWS patients with high and low tumor risks. It is also possible to predict whether patients are at risk of developing Wilms' tumor. Different screening protocols could therefore be offered to BWS patients, based on molecular diagnosis. In BWS patients with a telomeric defect (30% of cases), tumor management should involve abdominal ultrasound scans every 3 months, with clinical examination at alternate consultations, during the first 6 years of life. In BWS patients with a centromeric defect (70% of cases), tumor management should involve monthly clinical examinations during the first year, with a reference abdominal ultrasound scan at 3 months, followed by a clinical examination every 3 months for 6 years.

Plasma alpha-fetoprotein (AFP) levels have been put forward as a possible marker for routine tumor screening in children with BWS. AFP levels should be interpreted with a normal curve established specifically for BWS as AFP concentration is higher in patients with BWS than in healthy infants and children (Everman DB *et al.*, 2000; Clericuzio CL *et al.*, 2003).

Genetic counseling

Most of BWS are sporadic (85%) but about 15% BWS cases correspond to familial forms. The risk of recurrence in a family depends on the genetic cause of BWS in the proband.

Cytogenetic abnormality:

The risk to siblings of patients with BWS is up to 50% in case of a maternal 11p15 translocation or inversion. This risk is not clearly defined in BWS patients with an 11p15 duplication inherited from a father carrying a balanced translocation involving chromosome 11p15. **CDKN1C mutation:**

The risk to siblings of patients with BWS is up to 50% if the mother has the mutated *CDKN1C* gene. The children of a woman with a *CDKN1C* mutation have a 50% risk. The children of a man with a *CDKN1C* mutation have a theoretical risk of 0%, but with 50% of them will carry the mutation and the disease may be transmitted by girls to the next generation.

11p15 UPD:

The risk of recurrence is very low in cases of paternal UPD, as UPD results from a postzygotic event.

Épigenetic abnormalities:

Although rare, there are familial forms of BWS involving demethylation of *KCNQ10T1*, which are maternally transmitted. A microdeletion of the *KCNQ10T1* gene has been identified in one of these familial forms (Niemitz EL *et al.*, 2004). A few patients with hypermethylation of *H19* have also been shown to display a maternallyinherited microdeletion within *IC1* (Sparago A *et al.*, 2004). The recurrence risk for siblings and offspring of BWS patients with demethylation of *KCNQ10T1* or hypermethylation of *H19* is probably low. However, the frequency of microdeletions of imprinting centers is unknown and it is therefore difficult to generate an accurate figure for risk estimation.

Antenatal diagnosis

Prenatal diagnosis by ultrasound scan can be used to assess fetal growth and to detect abdominal wall defects, thereby helping to prevent neonatal complications.

Cytogenetic testing is appropriate for the diagnosis of translocation, inversion or duplication. Molecular diagnosis is also possible for 11p15 paternal UPD or *CDKN1C* gene mutation.

The reliability of testing for epigenetic modifications is unknown.

Beckwith-Wiedemann syndrome and assisted reproductive technology

Syndromes involving epigenetic alterations have recently been reported to occur in animals and humans conceived by ART. These include large offspring syndrome (LOS) in ruminants (Young L et al., 2001), BWS (DeBaun M et al., 2003; Gicquel C et al., 2003; Maher ER et al., 2003; Halliday J et al., 2004) and Angelman syndrome (Cox GF et al., 2002; Orstavik K et al., 2002) in humans. Various genetic and epigenetic mechanisms are involved in BWS and Angelman syndrome, but, following ART, the molecular defect in these imprinting disorders, always involves a loss of methylation of a maternallyimprinted methylated gene (demethylation of KvDMR1/KCNQ10T1 in BWS) suggesting that ART impairs the acquisition or maintenance of methylation marks on maternal imprinted genes. No specific procedure has yet been implicated in the epigenetic risk of ART-conceived patients. Indeed, ART-conceived Angelman syndrome and BWS patients were conceived by various procedures including classical IVF, ICSI, embryo cryopreservation, early or late embryo transfer, the use of various culture media (Cox GF et al., 2002; Orstavik K et al., 2002; DeBaun M et al.,

Gicquel C., Rossignol S., Le Bouc Y. Beckwith-Wiedemann Syndrome. Orphanet encyclopedia. March 2005. http://www.orpha.net/data/patho/GB/uk-BWS05.pdf 2003; Gicquel C *et al.*, 2003; Maher ER *et al.*, 2003; Halliday J *et al.*, 2004; Chang AS *et al.*, 2005). Large-scale and long-term outcome studies in children born as a result of ART should make it possible to estimate the exact risk of imprinting disorders after ART and to identify the underlying cause of this association.

References

Baujat G, Rio M, Rossignol S, Sanlaville D, Lyonnet S, Le Merrer M, Munnich A, Gicquel C, Cormier-Daire V, Colleaux L. Paradoxical NSD1 mutations in Beckwith-Wiedemann syndrome and 11p15 anomalies in Sotos syndrome. Am J Hum Genet. 2004; 74:715-20.

Beckwith JB. Nephrogenic rests and the pathogenesis of Wilms tumor: developmental and clinical considerations. Am J Med Genet. 1998; 79:268-73.

Bliek J, Maas SM, Ruijter JM, Hennekam RC, Alders M, Westerveld A, Mannens MM. Increased tumour risk for BWS patients correlates with aberrant H19 and not KCNQ10T1 methylation: occurrence of KCNQ10T1 hypomethylation in familial cases of BWS. Hum Mol Genet. 2001; 10:467-76.

Bliek J, Gicquel C, Maas S, Gaston V, Le Bouc Y, Mannens M. Epigenotyping as a tool for the prediction of tumor risk and tumor type in patients with Beckwith-Wiedemann syndrome (BWS). J Pediatr. 2004; 145:796-9.

Chang AS, Moley KH, Wangler M, Feinberg AP, Debaun MR. Association between Beckwith-Wiedemann syndrome and assisted reproductive technology: A case series of 19 patients. Fertil Steril. 2005; 83:349-54.

Cleary MA, van Raamsdonk CD, Levorse J, Zheng B, Bradley A, Tilghman SM. Disruption of an imprinted gene cluster by a targeted chromosomal translocation in mice. Nat Genet. 2001; 29:78-82.

Clericuzio CL, Chen E, McNeil DE, O'Connor T, Zackai EH, Medne L, Tomlinson G, DeBaun M. Serum alpha-fetoprotein screening for hepatoblastoma in children with Beckwith-Wiedemann syndrome or isolated hemihyperplasia. J Pediatr. 2003; 143:270-2.

Cox GF, Burger J, Lip V, Mau UA, Sperling K, Wu BL, Horsthemke B. Intracytoplasmic sperm injection may increase the risk of imprinting defects. Am J Hum Genet. 2002; 71:162-4.

DeBaun MR, Tucker MA. Risk of cancer during the first four years of life in children from The Beckwith-Wiedemann Syndrome Registry. J Pediatr. 1998; 132:398-400.

DeBaun MR, Niemitz EL, McNeil DE, Brandenburg SA, Lee MP, Feinberg AP. Epigenetic alterations of H19 and LIT1 distinguish patients with Beckwith-Wiedemann syndrome with cancer and birth defects. Am J Hum Genet. 2002; 70:604-11.

DeBaun M, Niemitz E, Feinberg A. Association of In Vitro Fertilization with Beckwith-Wiedemann Syndrome and Epigenetic Alterations of LIT1 and H19. Am J Hum Genet. 2003; 72:156-60.

Elliott M, Bayly R, Cole T, Temple IK, Maher ER. Clinical features and natural history of Beckwith-Wiedemann syndrome: presentation of 74 new cases. Clin Genet. 1994; 46:168-74.

Engel JR, Smallwood A, Harper A, Higgins MJ, Oshimura M, Reik W, Schofield PN, Maher ER. Epigenotype-phenotype correlations in Beckwith-Wiedemann syndrome. J Med Genet. 2000; 37:921-6.

Everman DB, Shuman C, Dzolganovski B, O'Riordan M A, Weksberg R, Robin NH. Serum alpha-fetoprotein levels in Beckwith-Wiedemann syndrome. J Pediatr. 2000; 137:123-7.

Fitzpatrick GV, Soloway PD, Higgins MJ. Regional loss of imprinting and growth deficiency in mice with a targeted deletion of KvDMR1. Nat Genet. 2002; 32:426-31.

Gaston V, Le Bouc Y, Soupre V, Burglen L, Donadieu J, Oro H, Audry G, Vazquez MP, Gicquel C. Analysis of the methylation status of the KCNQ1OT and H19 genes in leukocyte DNA for the diagnosis and prognosis of Beckwith-Wiedemann syndrome. Eur J Hum Genet. 2001; 9:409-18.

Gicquel C, Gaston V, Mandelbaum J, Siffroi J, Flahault A, Le Bouc Y. In vitro fertilization may increase the risk of Beckwith-Wiedemann syndrome related to the abnormal imprinting of the KCNQ1OT gene. Am J Hum Genet. 2003; 72:1338-41.

Grundy RG, Pritchard J, Baraitser M, Risdon A, Robards M. Perlman and Wiedemann-Beckwith syndromes: two distinct conditions associated with Wilms' tumour. Eur J Pediatr. 1992; 151:895-8.

Halliday J, Oke K, Breheny S, Algar E, Amor D. Beckwith-Wiedemann syndrome and IVF: a case-control study. Am J Hum Genet. 2004; 75:526-8.

Henneveld HT, van Lingen RA, Hamel BC, Stolte-Dijkstra I, van Essen AJ. Perlman syndrome: four additional cases and review. Am J Med Genet. 1999; 86:439-46.

Hoyme HE, Seaver LH, Jones KL, Procopio F, Crooks W, Feingold M. Isolated hemihyperplasia (hemihypertrophy): report of a prospective multicenter study of the incidence of neoplasia and review. Am J Med Genet. 1998; 79:274-8.

Itoh N, Becroft DM, Reeve AE, Morison IM. Proportion of cells with paternal 11p15 uniparental disomy correlates with organ enlargement in Wiedemann-beckwith syndrome. Am J Med Genet. 2000; 92:111-6.

Gicquel C., Rossignol S., Le Bouc Y. Beckwith-Wiedemann Syndrome. Orphanet encyclopedia. March 2005. http://www.orpha.net/data/patho/GB/uk-BWS05.pdf

Lam WW, Hatada I, Ohishi S, Mukai T, Joyce JA, Cole TR, Donnai D, Reik W, Schofield PN, Maher ER. Analysis of germline CDKN1C (p57KIP2) mutations in familial and sporadic Beckwith-Wiedemann syndrome (BWS) provides a novel genotype-phenotype correlation. J Med Genet. 1999; 36:518-23.

Lee MP, DeBaun M, Randhawa G, Reichard BA, Elledge SJ, Feinberg AP. Low frequency of p57KIP2 mutation in Beckwith-Wiedemann syndrome. Am J Hum Genet. 1997; 61:304-9.

Lee MP, DeBaun MR, Mitsuya K, Galonek HL, Brandenburg S, Oshimura M, Feinberg AP. Loss of imprinting of a paternally expressed transcript, with antisense orientation to KVLQT1, occurs frequently in Beckwith-Wiedemann syndrome and is independent of insulin-like growth factor II imprinting. Proc Natl Acad Sci U S A. 1999; 96:5203-8.

Maher ER, Reik W. Beckwith-Wiedemann syndrome: imprinting in clusters revisited. J Clin Invest. 2000; 105:247-52.

Maher ER, Brueton LA, Bowdin SC, Luharia A, Cooper W, Cole TR, Macdonald F, Sampson JR, Barratt CL, Reik W, Hawkins MM. Beckwith-Wiedemann syndrome and assisted reproduction technology (ART). J Med Genet. 2003; 40:62-4.

Morison IM, Becroft DM, Taniguchi T, Woods CG, Reeve AE. Somatic overgrowth associated with overexpression of insulin-like growth factor II. Nat Med. 1996; 2:311-6.

Neri G, Gurrieri F, Zanni G, Lin A. Clinical and molecular aspects of the Simpson-Golabi-Behmel syndrome. Am J Med Genet. 1998; 79:279-83.

Niemitz EL, DeBaun MR, Fallon J, Murakami K, Kugoh H, Oshimura M, Feinberg AP. Microdeletion of LIT1 in familial Beckwith-Wiedemann syndrome. Am J Hum Genet. 2004; 75:844-9.

Orstavik K, Eiklid K, Van der Hagen C, Spetalen S, Kierulf K, Skjeldal O, Buiting K. Another case of imprinting defect in a girl with Angelman syndrome who was conceived by intracytoplasmic sperm injection. Am J Hum Genet. 2002; 72:218-9.

Pettenati MJ, Haines JL, Higgins RR, Wappner RS, Palmer CG, Weaver DD. Wiedemann-Beckwith syndrome: presentation of clinical and cytogenetic data on 22 new cases and review of the literature. Hum Genet. 1986; 74:143-54.

Pilia G, Hughes-Benzie RM, MacKenzie A, Baybayan P, Chen EY, Huber R, Neri G, Cao A, Forabosco A, Schlessinger D. Mutations in GPC3, a glypican gene, cause the Simpson-Golabi-Behmel overgrowth syndrome. Nat Genet. 1996; 12:241-7.

Reik W, Walter J. Genomic imprinting: parental influence on the genome. Nat Rev Genet. 2001; 2:21-32.

Schneid S, Vazquez M, Vacher C, Gourmelen M, Cabrol S, Le Bouc Y. The Beckwith-Wiedemann syndrome phenotype and the risk of cancer. Med Pediatr Oncol. 1997; 28:411-5.

Smilinich NJ, Day CD, Fitzpatrick GV, Caldwell GM, Lossie AC, Cooper PR, Smallwood AC, Joyce JA, Schofield PN, Reik W, Nicholls RD, Weksberg R, Driscoll DJ, Maher ER, Shows TB, Higgins MJ. A maternally methylated CpG island in KvLQT1 is associated with an antisense paternal transcript and loss of imprinting in Beckwith-Wiedemann syndrome. Proc Natl Acad Sci U S A. 1999; 96:8064-9.

Sotelo-Avila C, Gonzalez-Crussi F, Fowler JW. Complete and incomplete forms of Beckwith-Wiedemann syndrome: their oncogenic potential. J Pediatr. 1980; 96:47-50.

Sparago A, Cerrato F, Vernucci M, Ferrero GB, Silengo MC, Riccio A. Microdeletions in the human H19 DMR result in loss of IGF2 imprinting and Beckwith-Wiedemann syndrome. Nat Genet. 2004; 36:958-60.

Thorburn MJ, Wright ES, Miller CG, Smith-Read EH. Exomphalos-macroglossia-gigantism syndrome in Jamaican infants. Am J Dis Child. 1970; 119:316-21.

Weksberg R, Nishikawa J, Caluseriu O, Fei YL, Shuman C, Wei C, Steele L, Cameron J, Smith A, Ambus I, Li M, Ray PN, Sadowski P, Squire J. Tumor development in the Beckwith-Wiedemann syndrome is associated with a variety of constitutional molecular 11p15 alterations including imprinting defects of KCNQ1OT1. Hum Mol Genet. 2001; 10:2989-3000.

Wiedemann HR. Familial malformation complex with umbilical hernia and macroglossia - A "new syndrome"? J Genet Hum. 1964; 13:223-32.

Young L, Fernandes K, McEvoy T, Butterwith S, Gutierrez C, Carolan C, Broadbent P, Robinson J, Wilmut I, Sinclair K. Epigenetic change in IGF2R is associated with fetal overgrowth after sheep embryo culture. Nat Genet. 2001; 27:153-4.